

Extraction and Characterization of “Batuan” [*Garcinia binucao* (Blco.) Choisy] Seed Protein

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ABSTRACT

“Batuan” [*Garcinia binucao* (Blco.) Choisy], an indigenous, lesser known member of the Gutierrezia family with export potential is underutilized and understudied. The present study was carried out to extract and characterize the protein in “batuan” [*Garcinia binucao* (Blco.) Choisy] seeds for nutritional quality assessment. Protein content of “batuan” seed meal was $8.9 \pm 0.59\%$ dry basis. Solubility fractionation of “batuan” seed meal showed globulin and glutelin as the major seed proteins. SDS-PAGE resolved the globulin and glutelin into three groups of polypeptides with molecular weights of about 20 – 54 kDa. Amino acid analysis revealed that seed protein contained all the essential amino acids with leucine as the most abundant while tryptophan, the least. “Batuan” seed proteins were mostly made up of acidic and hydrophobic amino acids with glutamic acid (2.67%) as the highest. Nutritional assessments including E/T (38.4%), amino acid score (1.6%), predicted PER (3.2-3.7) and estimated BV (98.3%) suggested that the seed proteins are of good quality. Hence, “batuan” seeds has a promising potential as an important sources of valuable proteins and amino acids for use as food supplement/enhancing ingredient.

Keywords: amino acid, “batuan” seed protein, nutritional, tryptophan

INTRODUCTION

Seed storage proteins which accumulate in high amounts in some plant seeds as sources of nutrients for germinating seeds constitute an important source of dietary proteins for human consumption. In the Philippines, many edible, indigenous, seed-bearing fruit trees which are known only by the local people provide income as well as cheap sources of their dietary proteins. “Batuan”, *Garcinia binucao* (Blco.) Choisy, is one of the lesser-known, indigenous, seed-bearing fruit trees identified with export potential (de la Cruz, 2013). The fruit which is mainly used as souring agent in native dishes contains several (more or less 7) large, edible seeds (Quevedo, 2013; Florido and Cortiguerra, 2003). However, the quality and quantity of the seed protein of “batuan” fruit (Figure 1) remains little if none at all has been published to date. Quevedo (2013) revealed that the protein content of “batuan” fruits is generally low (less than 10%) with the seeds containing the highest.

Characterization of the seed storage proteins in “batuan” fruit is important since it is consumed and considered as a favorite souring agent by Filipinos especially in some part of the provinces of Negros Occidental, Iloilo, Leyte, Masbate and in other places in the Philippines (Florido and Cortiguerra, 2003; Cojuangco, 2012; de la Cruz, 2013). During fruiting season, the fruits are processed into various products such as salted puree, jam, jelly, prunes, and candies. Thus, information in terms of protein quality and quantity are very valuable and may make significant contribution to its nutrient quality and processing properties as well as its potential application in food processing. Furthermore, analysis of the seed storage proteins in “batuan” may also provide insight into characterization of genetic diversity, improvement of nutritional quality, and seed development for “batuan”.

This article presents the results on the extraction and characterization of the protein in “batuan” seed. “Batuan” seed proteins were evaluated for their relative amount and amino acid composition. An attempt was also made to predict its nutritional quality.



Figure 1. A. The “batuan” [*G. binucao* (Blco.) Choisy] fruits. B. Cross-section of “batuan” fruit with seeds.

MATERIALS AND METHODS

Materials

Seeds of “batuan” were obtained from harvested mature fruits at the Pomology Project of the Visayas State University, Visca, Baybay City, Leyte. The fruits were authenticated from the University of the Philippines Los Baños (UPLB) Museum of Natural History, College, Laguna, Philippines. Analytical grade acrylamide, ammonium persulfate (APS), bis-acrylamide, molecular weight markers 14,400-200,000 Da (BIORAD), bromphenol blue, Coomassie Brilliant Blue G-250, dihydrate hydrogen phosphate, glycine, potassium dihydrogen phosphate, sodium chloride, sodium dodecyl sulfate (SDS), sodium hydroxide, and Trizma base, acetic acid, Bradford reagent, ethanol, hydrochloric acid, beta-mercaptoethanol, N,N,N',N'-tetramethylethylenediamine (TEMED) were procured from commercial suppliers.

The dialysis tubing cellulose membrane (MWCO 12,000) was purchased from Sigma Chemicals.

Preparation of the Defatted “Batuan” Seed Meal

Dried “batuan” seed endosperm was ground in a blender, then passed through a 40-mesh sieve and defatted using direct solvent (hexane)

method with constant stirring at room temperature under the fume hood for 24 hr. The meal was stored at 5°C after removal of hexane and air-drying under the fume hood.

Protein Fractionation

Storage proteins from the defatted “batuan” seed meal were extracted with stirring at 4°C for 1 hr in 1:20 w/v water, 0.5 M NaCl, 70% ethanol and 0.1 M NaOH according to the Osborne solubility procedure (Osborne, 1924). All extractions were carried out twice. Albumin, globulin and glutelin fractions were dialyzed. All protein fractions were freeze-dried, then the dried samples were stored at 5°C for subsequent use.

Preparation of Total “Batuan” Seed Protein Extract

Total “batuan” seed protein was obtained from defatted seed meal by dispersing at 1:2 w/v in 20 mM Tris-HCl buffer. The mixture was stirred for 2 h at 4°C, centrifuged at 10,000 rpm for 10 min at 4°C and filtered through Whatman No. 4 filter paper. The supernatant was freeze-dried and the dried sample extract was stored at 4°C for subsequent use.

Protein Content

Protein contents in the extract and the Osborne fractions were assayed by Bradford's method (Bradford, 1976) using bovine serum albumin (BSA) as protein standard. Protein concentrations of the samples were calculated using the linear regression formula from the bovine serum albumin (BSA) standard curve.

SDS-PAGE Analysis

The estimated molecular weights of the polypeptides in the different protein solutions (crude extract and Osborne protein fractions) were determined by the sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) using 12.5% polyacrylamide gel according to Laemmli (1970) and low molecular weight protein standard markers (BIORAD). The lyophilized crude extract and Osborne fractions were dissolved in 1 mL of 20 mM Tris-HCl buffer at pH 8.8. The solution was then centrifuged at 10,000 rpm for 2 min at 4°C to obtain the purified

sample. Aliquots of the extract and Osborne fractions were added with sample buffer, heated for 5 min and cooled to room temperature before loading to the wells at a concentration of $0.25 \text{ mg } \mu\text{L}^{-1}$. Coomassie brilliant blue R-250 was used for staining.

Amino Acid Analysis

Dried, powdered, defatted seed endosperm meal sample was sent for amino acid content analysis at the SGS Thailand, Inc. and analyzed following their in-house method based on the protocol for amino acid analysis in the Handbook of Food Analysis Vol. 1 and using Gas Chromatography-Mass Spectrophotometry.

Evaluation of Nutritional Parameters

The amino acid composition was used to estimate the nutritional value of the protein based on the provisional amino acid scoring pattern for preschool children (WHO, 2007). The parameters such as the proportion of essential amino acids with total amino acids in the protein (E/T %), amino acid score or chemical score, predicted protein efficiency ratio (PER) and estimated biological value (BV) were used for nutritional quality assessment. PER values were estimated using the regression equations proposed earlier by Alsmeyer *et al.* (1974) which are outlined as follows:

$$\begin{aligned} PER I &= -0.684 + 0.456(Leu) - 0.047(Pro) \\ PER II &= -0.468 + 0.454(Leu) - 0.105(Tyr) \\ PER III &= -1.816 + 0.435(Met) + 0.780(Leu) + 0.211(His) - 0.944(Tyr) \end{aligned}$$

BV of protein in “batuan” seed was estimated using the equation developed by Mørup and Olesen (1976) and is shown below:

$$BV = 10^{2.15} \times q_{lys}^{0.141} \times q_{phe+tyr}^{0.60} \times q_{met+lys}^{0.77} \times q_{thr}^{2.14} \times q_{trp}^{0.21}$$

where:

$$q = \frac{ai \text{ sample}}{ai \text{ reference}} \text{ for } ai \text{ sample} \leq ai \text{ reference}$$

or

$$q = \frac{ai \text{ reference}}{ai \text{ sample}} \text{ for } ai \text{ sample} \geq ai \text{ reference}$$

ai = mg of amino acid per g of total essential amino acids.

Statistical Analysis

Protein content and SDS-PAGE analyses were done in triplicates while amino acid analysis in duplicate. Results on protein content were reported as mean \pm SD.

RESULTS AND DISCUSSION

Fractionation of "Batuan" Seed Proteins

The crude protein of "batuan" seed endosperm on moisture free basis was $8.9 \pm 0.52\%$. Osborne's protein fractionation procedure extracted 92.82% of the proteins (Table 1) and contained albumin, globulin, prolamin and glutelin. Globulins are the most abundant, accounting for about 67.49% of the total soluble proteins followed by glutelin (20.40%), albumin (3.81%) and prolamin (1.12%). This finding is similar to those of Moringa seeds where globulins and glutelins constitute the major storage proteins (Reyes, 2010). Furthermore, there are no previously reported data on the seed storage proteins of "batuan" in the Philippines and from other countries to compare the results of this present work.

Table 1. Seed storage proteins (%) of "batuan" fractionated using the Osborne solubility scheme.

	PROTEIN CONTENT (% db) ¹	% OF TOTAL PROTEIN CONTENT
Initial Protein	8.92 \pm 0.59	
Total soluble protein	8.28 \pm 0.61	92.82
Albumin	0.34 \pm 0.02 ^c	3.81
Globulin	6.02 \pm 0.05 ^c	67.49
Prolamin	0.10 \pm 0.00 ^a	1.12
Glutelin	1.82 \pm 0.54 ^b	20.4
Nonsolubilized residue	0.025 \pm 0.00	0.28
Total crude protein recovered	8.305 \pm 0.61	93.1

¹Average of 3 trials

Means within a column followed by a common superscript are not significantly different at 5% level, LSD.

Molecular Weight Determination by SDS-PAGE

The different Osborne fractions from the defatted "batuan" seed meal were further resolved on SDS-PAGE and the electrophoretogram is shown

in Figure 2. Globulin fractions revealed polypeptides of low molecular weights at approximately 51.85, 27.52 and 20.97 kDa. The glutelin fractions also exhibited three prominent polypeptide bands with molecular weights of about 53.92, 27.92 and 21.06 kDa.

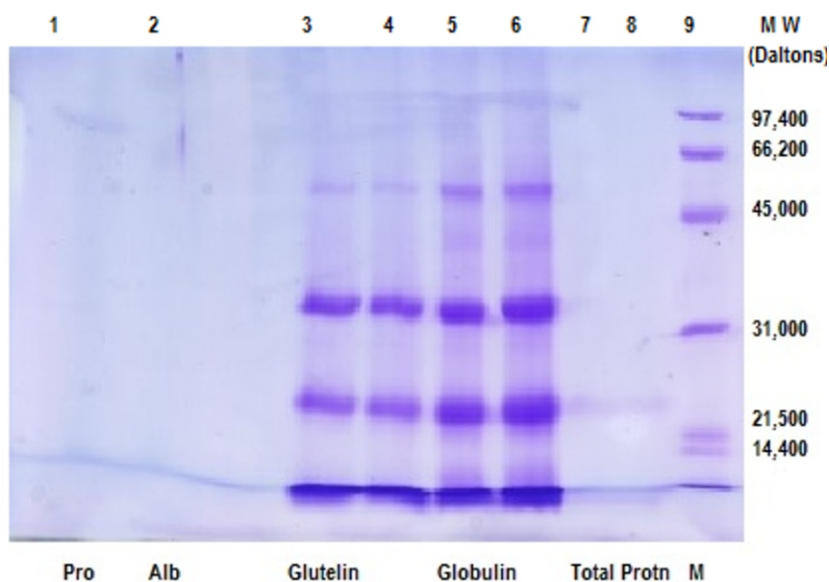


Figure 2. SDS-PAGE of the seed storage protein fractions of “batuan”.
 Lane: 1- prolamine; Lane 2 – albumin; Lanes 3 & 4 – glutelins;
 Lanes 5 & 6 – globulin; Lanes 7 & 8 – total protein;
 Lane 9 – protein standard

Meanwhile, no bands were observed in the albumin, and prolamin fractions. This is probably due to the low protein content in the albumin or prolamin fractions. However, it is also possible that the protein content for albumin and prolamin obtained using Bradford assay was an estimate for other compounds such as polyphenols and some alkaloids that were also extracted by the solvents from the seed matrix. These impurities when present in high concentration in the protein sample may have interfered with the assay by associating with the Coomassie dye, giving a response at 595 nm. Compton and Jones (1985) pointed out that bases, detergents and other non-protein compounds when present at high concentration in the protein sample interfere with Bradford assay, resulting in an over estimation of the protein content.

The total protein extract exhibited less prominent polypeptide band of approximately 21.32 kDa identical to the globulin and glutelin bands. The

result implies that the separation gel buffer (20 mM Tris-HCl) used in the extraction of the total proteins in defatted “batuan” seed meal did not efficiently solubilize most of the proteins present. Probably, the concentration of Tris-HCl in the separation buffer (20 mM) was not enough to isolate the soluble proteins in the endosperm. Tris-HCl was used as the extractant of the total proteins in the defatted “batuan” seed meal since it is a component of the separation gel of the SDS-PAGE, and has been one of the widely used extraction buffer in protein purification as well as in the extraction of nucleic acids.

Globulins, the most widely distributed groups of storage proteins are oligomeric proteins consisting of 2 or more subunits that can be separated under mild dissociating conditions. Bewly and Black (1994) pointed out that these subunits are composed of a number of polypeptide chains containing variable amino acid sequence between homologous subunits. Based on their sedimentation coefficients, globulins are classified as being the 7S vicilin-type and the 11S legumin-type which exhibit considerable variation in their structure resulting from post translational processing.

In this study, both the globulin and the glutelin fractions contained three groups of polypeptides with almost similar molecular weights. However, the 53.92 kDa glutelin band was not very visible compared to the 51.85 kDa globulin band which may be due to a lower concentration of glutelin compared to globulin. Analysis of the 21.06 kDa of the glutelin band showed similarity in the molecular weight with the 19 – 22 kDa basic subunit of rice glutelins (Robert *et al.*, 1985; Kagawa *et al.*, 1998). Several earlier studies reported that glutelins showing variations in structures and molecular weights are also structurally similar to the 11S globulins (Zhao *et al.*, 1983; Wen and Luthe, 1985). These observations may also explain the similarity of the number of glutelin and globulin polypeptides obtained in this study. The identities as well as the characteristics of these low molecular weight polypeptides in “batuan” seeds have never been published to date. Thus, more detailed studies to give insights on its characteristics and properties are needed.

Amino Acids Profile

The amino acids composition of the “batuan” seed endosperm is shown in Table 2. Glutamic acid (2.67% dry basis) was found as the most abundant amino acid of the “batuan” seed proteins followed by aspartic acid (0.93%) and leucine (0.90%). This finding is similar with those

obtained in its close relative, mangosteen, *G. mangostana* (Ajayi *et al.*, 2013) as well as to other fruit seeds such as tamarind (Glew *et al.* (1997), and pigeon proteins (Etonihu *et al.*, 2009), with glutamic acid contents of 10.938 g 100g⁻¹, 2.82 mg g⁻¹, 14.64 g 100 g⁻¹ (dry weight), respectively.

Moreover, the total combined amount (3.6%) of acidic amino acids was higher than those of the uncharged polar amino acids (1.36%) and the basic amino acids (0.48%) and is almost the same as that of the hydrophobic amino acids (3.5%). This suggests that the proteins in "batuan" seeds and their protein fractions may be of acidic nature. Moreover, the acidic and hydrophobic amino acids in "batuan" seeds are almost equal in proportion. The results suggest that the lower molecular weight polypeptides in "batuan" seeds may have amphiphatic character, having both a hydrophobic region interacting with lipids and a positively charged hydrophilic region interacting with water and other negatively charged residues.

Table 2. Amino acid composition (% dry basis) of "batuan" [*G. binucao* (Blco.) Choisy] seeds and the percentage (%) essential amino acid requirements for preschool children (3-10 years) (WHO, 2007)

AMINO ACID	BATUAN SEED PROTEIN (%) ¹	WHO (2007) (%)
Valine	0.52	2.9
Leucine	0.90	4.4
Isoleucine	0.43	2.3
Threonine	0.37	1.8
Methionine	0.15	1.8
Phenylalanine	0.47	3.0
Lysine	0.27	3.5
Histidine	0.21	1.2
Tryptophan	0.11	4.8
Total essential amino acids (E)	3.43	20.9
Alanine	0.43	
Glycine	0.43	
Serine	0.25	
Proline	0.48	
Aspartic acid	0.93	
Glutamic acid	2.67	
Tyrosine	0.27	
Cystine	0.04	
Total nonessential amino acids (N)	5.50	
Total amino acids	8.18	
Classified distribution of amino acids		
1. Hydrophobic	3.51	
2. Uncharged polar	1.36	
3. Basic	0.48	
4. Acidic	3.60	

¹ Average of 2 measurements

Murray and Roxburgh (1984) as reported by Tounkara et al. (2013) also revealed that high levels of albumin elevated the sulfur-containing amino acids in chickpea proteins. In the present study, the amount of sulfur-containing amino acids which are most likely associated with albumins were low, supporting the observation of the absence or the presence of trace amount of albumin in “batuan” seed proteins.

Table 2 further shows that “batuan” seed endosperm contains nine essential amino acids with a total combined amount of 3.43% dry basis. Leucine (0.90%) was the most abundant followed by valine (0.52%) and phenylalanine (0.47%). However, the total essential amino acids in “batuan” seed protein (3.43%) was lower than the 20.9% total essential amino acid requirement for preschool children (WHO, 2007) but higher compared to the reported 0.58% essential amino acids in tamarind seed (Glew et al., 1997). Nonetheless, “batuan” seeds could supply 16.4% total essential amino acids, 10.6% total S-containing amino acids and 24.72% total aromatic amino acids of the reference values as reported by WHO (2007). As pointed out by Krzysciak (2011), the aromatic amino acids, beside their structural function in proteins, are precursors of many important biological compounds including neurotransmitters and hormones in the human body. Atmaon (2004) also reported that the S-containing amino acids cysteine and methionine are known to possess antioxidant property which is influenced by the type of oxidant stress and the physiological conditions. In addition, leucine, the only dietary amino acid with the capability of stimulating muscle synthesis (Etzel, 2004), having the highest value (0.9%) among the essential amino acids in “batuan” seed, could also deliver 20.4% of required leucine. Thus, the amount of essential amino acids present in “batuan” seeds would be a significant value addition or contribution to its nutritional quality as well as a good complementary source of dietary proteins. At present, there are no previously reported data on the amino acid content of “batuan” seed proteins in the Philippines and from other countries with which the results of this present work can be compared.

Estimation of Nutritional Quality and Quantity

The nutritional quality of protein is principally determined by its amino acids composition. Certain nutritional parameters were quantified using the amino acid composition (Table 2).

In 1973, the World Health Organizations had suggested the

provisional amino acid scoring pattern for an ideal protein. The pattern recommends that for a good protein, the E/T ratio should be at least 36%. “Batuan” seed protein exceeded this criterion suggesting that the proteins of “batuan” seed are of good quality. Moreover, the obtained E/T ratio for “batuan” seed proteins is higher compared to the 30.54% for cassava tuber-associated proteins obtained by Azucena (2005).

Computation of the predicted PER and estimated BV for “batuan” seed protein based on their amino acid composition was very satisfactory. It is clear from the calculated values in Table 3 that the equations developed by Alsmeyer et al. (1974) for predicting PER and by Mørup and Olesen (1976) for estimating BV, originally designed for meat products, were applicable to “batuan” seed proteins. In general, the protein efficiency ratio (PER) below 1.5 implies a protein of low or poor quality, while between 1.5 and 2.0 indicates an intermediate protein quality and above 2.0 means of high quality protein (Friedman, 1996). The predicted PER values for “batuan” seed protein calculated using PER equations I to III (3.2 to 3.7) as shown in Table 3, were in the range of high quality protein and relatively comparable with the reported PER of 3.1 for whole egg, & higher than the 2.9 for beef, 2.5 for casein & cow's milk, 2.2 for soy protein, & 0.8 for wheat gluten (Hoffman & Falvo, 2004). This implies that the “batuan” seed protein is of better quality.

Table 3. Nutritional evaluation of "batuan" seed proteins.

NUTRITIONAL PARAMETER	TOTAL "BATUAN" SEED PROTEIN
E/T, % ¹	38.4
Amino acids score (%)	
Tryptophan	1.7
Lysine	5.6
Methionine + Cysteine	6.2
Phenylalanine + Tyrosine	11.5
Leucine	14.7
Valine	13.0
Isoleucine	18.7
Histidine	32.5
Predicted Protein Efficiency Ratio	
PER I	3.2
PER II	3.3
PER II	3.7
Estimated Biological Value	98.23

¹E/T, % = proportion of essential amino acids and total amino acids. Amino acid scores were evaluated using the reported essential amino acid pattern as reference (WHO, 2007).

The estimated biological value (BV) of “batuan” seed protein was lower compared to the reported 100% of egg, but higher than the 93% of milk, 75% of beef & fish, 72% of corn (Whitney & Flores, 2010), 74% of soy protein, & 64% of wheat gluten (Hoffman & Falvo, 2004). The high estimated BV of the protein of “batuan” seed suggests that the protein would be efficiently utilized by the body. However, no comparison could be made on the predicted BV of the “batuan” seed protein in other places in the Philippines and other countries due to lack of published studies on nutrient evaluation.

CONCLUSION

Globulins (67.49%) and glutelins (20.40%) were the major seeds storage proteins of “batuan” based on Osborne fractionation scheme and Bradford assay. Three groups of polypeptides in the Osborne's globulin and glutelin fractions with molecular weights ranging from 20 – 54 kDa were resolved on SDS-PAGE. Glutamic acid (2.67%) was the most abundant amino acid in “batuan” seed protein followed by aspartic acid (0.93%) and leucine (0.9%). “Batuan” seed proteins contained more of the acidic amino acid groups (3.60%) and the hydrophobic amino acids (3.51%) than the uncharged polar (1.36%) and basic amino acids (0.48%). Furthermore, all essential amino acids were present in “batuan” seed protein and leucine (0.90%) was the most abundant. Ratio of essential to total amino acids (38.4%) exceeded the 36% criterion suggested by FAO/WHO/UNO (1975) for good quality protein. Based on amino acid score, tryptophan (1.67%) was the most deficient essential amino acid. Predicted PER (3.2-3.7) of “batuan” seed protein is within the range for efficiently absorbed and utilized proteins. Estimated BV (98.3%) was also high. Nutritional assessments suggested that “batuan” seed proteins are of good quality and could be a cheap natural source of supplementary dietary protein.

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