

Heavy metal tolerance of filamentous fungi from the sediments of Visayas State University wastewater pond

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ABSTRACT

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The ability to tolerate high concentrations of heavy metals is one important characteristic of organisms that can be used for bioremediation. In this study, the heavy metal tolerance of filamentous fungi isolated from the VSU wastewater settlement pond was investigated. Specifically, the research was done to determine the Cd, Cu, Fe and Zn in the sediments of the pond, isolate and identify filamentous fungi from these sediments, identify the most tolerant isolate, and determine the minimum inhibitory concentration of heavy metals to the identified isolate. Isolation of filamentous fungi from a composite of eight sediment samples was done through serial dilution and plating using Potato Dextrose Agar (PDA). Isolates were then purified using Sabouraud Dextrose Agar (SDA). Colony and microscopic characteristics of the isolates were used to identify the isolates to genus level only. The tolerance of the isolates to Cd, Cu, Fe, and Zn were then compared through analysis of their tolerance index based on the colony extension radius. The Minimum Inhibitory Concentrations (MIC) of these heavy metals were obtained for the isolate with the highest tolerance index. The Cd, Cu, Fe, and Zn contents of the composite sediment sample were <0.0002mg kg⁻¹, 0.0203mg kg⁻¹, 7.419mg kg⁻¹, and 0.106mg kg⁻¹, respectively. Three filamentous fungi were successfully isolated from the sediments and were identified as Rhizopus sp., Mucor sp. and Trichoderma sp. Among these isolates, Rhizopus sp. was the most tolerant to all the heavy metals tested. Average range of MIC values of Cd, Cu, Fe and Zn to Rhizopus sp. were determined at 5mM<MIC≤6.5mM, 10mM<MIC≤15mM, 30mM<MIC≤35mM, and 25mM<MIC≤30mM, respectively. These results showed that the Rhizopus sp. isolated is the most potential candidate in bioremediating heavy metal contaminated areas.

Keywords: Industrial effluents, heavy metal pollution, mycoremediation, tolerance index, filamentous fungi

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INTRODUCTION

Heavy metal pollutants are found in various wastewaters discharged from industrial plants, business centers, and other establishments especially those that are near human settlements (Tchounwou 2014). When untreated, these effluents cause serious environmental stress and damage to the surrounding communities and ecosystems. They can cause fish kills (Slaninova et al 2014), poisoning (Rholand 2013), and various diseases (Jarup 2003). Thus, wastewater treatment is essential, and treatment through bioremediation is an excellent, safe, economical, and ecologically-friendly option.

Bioremediation is a method used to provide techniques in cleaning up pollution by enhancing the same biodegradation process that occurs in nature through the use of biological agents (Rhodes 2013). It can be achieved through either microbial remediation (using microorganisms) or phytoremediation (using plants) (Kensa 1970). Microbial remediation is the natural means of bioremediation since it involves the decomposers which occupy the bottom of the food chain, responsible for the decomposition of waste matter (Rhodes 2012, Schmitz 2008).

Among the known bioremediators are the filamentous fungi. They have hyphae, which are long, tubular structures that have rigid walls while some are reinforced by perforated cross-walls called septae (Watkinson et al 2015). Physiologically, the hyphae serve to absorb nutrients from the environment as well as translocate contaminants in the same manner they translocate nutrients from organic matter (Brand 2012, Harms et al 2011). This ability enables them to tolerate high concentrations of heavy metals in the environment and become effective for bioremediation (Bhattacharya 2011).

Heavy metal tolerance is the level at which an organism can tolerate high doses of heavy metals due to the organism's physiological and biochemical adaptation to these substances (Singh et al 2011). This characteristic is an important requirement for an organism to participate in bioremediation of toxic metals because without this, these microorganisms die when subjected to contaminated areas resulting to inefficient bioremediation (Ezzourhi et al 2009).

Heavy metal tolerant filamentous fungi are potent tools in bioremediation, for they can facilitate the removal of toxic heavy metals through translocation of contaminants in the same manner as they take in the nutrients from organic matter (Harms et al 2011). As these fungi are studied further, they may help prevent the loss of our natural resources such as food in the case of fish kills (Slaninova et al 2014), cleaning our environment and making a better environment for future generations.

The main campus of Visayas State University (VSU) is composed of upper and lower campuses. In the lower campus, the wastewater treatment is held in a wastewater pond. The pond is the reservoir of all waste effluents from students' dormitories, laboratories, faculty and staff housing units, academic buildings, hostel, hospital, and other structures inside the university. This pond treats the pollutants by sedimentation process wherein solid pollutants settle to the bottom.

Thus, this study aimed to determine the Cd, Cu, Zn, and Fe concentrations in the sediments of the VSU wastewater pond, isolate and identify filamentous fungi from these sediments, select the most tolerant isolate, and determine the minimum inhibitory concentration of these heavy metals to the isolate.

MATERIALS AND METHODS

Sampling and Preparation

All materials including the culture media were sterilized at 121°C, 15psi for 15min. Sediment samples were collected from eight (8) different points in the sampling site (Figure 1) using a grab sampler. A 500 gram-composite sample was placed in a sterile bottle (Ezzouhri et al 2009), and the remaining samples were airdried and sent to the F.A.S.T. Laboratory in Mandaue City, Cebu, Philippines for analysis of Cd, Cu, Fe, and Zn. Data for the heavy metal content of the VSU wastewater pond in 2012 and 2016 were taken from the unpublished research of Valenzona and Dacera et al, respectively.



Figure 1. Sampling points within Visayas State University wastewater pond. Source: Google Maps 2019

Isolation of Filamentous Fungi

One (1) g of the natural composite samples was suspended in a sterile 500mL beaker filled with 100mL distilled water and then covered with parafilm. The mixture was agitated for 20min at room temperature. Subsequently, the mixture was diluted in sterile beakers at 10, 100, 1000, and 10000 fold of serial dilution (Ezzouhri et al 2009).

Thirty-nine (39) g of Potato Dextrose Agar (PDA) were dissolved in an Erlenmeyer flask containing 1L distilled water and covered with an autoclavable plastic. The solution was then sterilized at 121°C, 15psi for 15min prior to isolation of the heavy metal- tolerant fungi. Three replicates of 100μ L aliquots from the

different dilutions of the sample sediments were plated onto the sterile plates containing solidified PDA, using lawn method. The inoculated plates were incubated for three days at 25°C. The distinct colonies that grew on the PDA plates were picked by slicing-off the agar from the plate and transferring them onto fresh SDA medium. They were then incubated for three days at 25°C (Ezzouhri et al 2009). Subsequent transfers to fresh Sabouraud Dextrose Agar (SDA) plates were made until pure cultures of the isolates were attained.

Identification of the Filamentous Fungal Isolates

Colonial and cellular characteristics of the filamentous fungal isolates were evaluated to identify them down to their lowest possible taxon. These characteristics include texture and color as viewed under the stereoscope.

For cellular characterization, the fungal colony of each isolate was inoculated on a sterile glass slide with PDA and placed inside a petri plate with sterile wet cotton to provide moisture. After at least three days of incubation, it was stained with lactophenol cotton blue and topped with coverslip. When the fungal colonies were already sufficiently stained, they were examined under a compound light microscope (True Vision Microscope, USA) to assess their cellular characteristics. Photomicrographs were taken using Amscope 5MP camera directly mounted on the microscope to aid in identification.

Screening and Selection of Isolates with High Heavy Metal Tolerance

The filamentous fungal isolates were screened for their tolerance to cadmium (Cd), copper (Cu), iron (Fe), and zinc (Zn) based on the method of Ezzouhri et al (2009). Each of the PDA plates was supplemented with 1mM of a specific metal salt. Aseptically, disks of mycelium were inoculated onto the PDA plates and incubated at 25°C. After 7 days incubation, fungal growth was determined by measuring the radius (mm) of the colony extension of the fungal mycelia in the agar. Tolerance index was then calculated as the ratio of the extension radius of the treated colony (with heavy metal) to the untreated colony (without heavy metal). The tolerance indices of the isolates were analyzed statistically identify the isolate showing the highest tolerance to all the heavy metals tested.

Minimum Inhibitory Concentration (MIC) Testing

Testing for the MIC was carried out only on the most tolerant filamentous fungal isolate. Each of the fresh SDA media was first supplemented with a specific heavy metal based on the concentrations used by Ezzouhri et al (2009), 0 to 35mM or higher until the concentration that totally inhibited the growth of the fungus was reached. The media containing different concentrations of the heavy metal salts were inoculated with 6mm agar disks from the young fungal colonies. The inoculated plates, in triplicate, were incubated for 7 days at 25°C. The minimum concentration of the heavy metal that inhibited visible growth of the isolate was regarded as the MIC (Ezzouhri et al 2009).

Experimental Design and Statistical Analysis

The experiment was laid in a completely randomized design (CRD). Analysis of variance (ANOVA) was performed followed by Tukey's Honest Significant Difference (HSD) test for multiple comparison of means at alpha=0.01%. All the analyses were carried out using IBM SPSS Statistics 22 software.

RESULTS AND DISCUSSION

Heavy Metal Content of the Sediments from the VSU Wastewater Pond

The VSU wastewater pond is the wastewater treatment facility of the university where effluents from faculty and staff housing, academic buildings, and laboratories are treated by sedimentation process. The heavy metal contents of the pond in 2012, 2016, and 2019 are shown in Table 1. The concentration of Cd and Cu from 2012 (Valenzona 2012) to 2016 (Dacera et al 2016) were from 0.074mg kg⁻¹ to 0.83mg kg⁻¹, 0.0324mg kg⁻¹ to 78.3mg kg⁻¹ and 0.9703 to 25mg kg⁻¹, respectively. This probably means that there was a continuous heavy metal discharge into the pond from 2012 to 2016, which resulted to the increase in the heavy metal concentrations in the pond. However, in 2019, the concentration of heavy metals decreased. There could have been a decrease in the heavy metal input to the pond or probably there was bioremediation carried out by the microbial population and plant species found in the area, or a combination of both. In 2012 and 2016, the studies of Valenzona and Dacera, respectively, showed that the area was devoid of vegetation, but in 2019, Eliseo reported that plants were already found growing around the area.

Heavy Metal		Concentration (mg per kg)	
	Valenzona 2012	Dacera et al 2016	Eliseo 2019
Cadmium	0.0747	Less than 0.83	Less than 0.0002
Copper	0.0324	78.3	0.0203
Lead	0.9703	25	0.00315
Iron	No data	No data	7.419
Zinc	No data	No data	0.106

 $Table\,1.\,Heavy\,metal\,contents\,of\,the\,VSU\,was tewater\,pond\,sediments\,in\,2012,2016,and\,2019$

Characteristics of the Filamentous Fungal Isolates

Figure 2 shows the colony and cellular characteristics of the three filamentous fungi isolated in this study. The fungi isolated from the sediments designated as Isolate 1, Isolate 2 and Isolate 3 belong to the genera *Rhizopus, Mucor* and *Trichoderma*, respectively. Isolate 1 has black-colored colonies producing circular sporangia enclosing the spores, and it had non-septate stolons (Figure 2 first row). These morphological characteristics describe the genus *Rhizopus* of Phylum Oomycota. The genus *Aspergillus* of Phylum Eumycota may also have some of these characteristics, except that they produce septate mycelia.



Figure 2. Colony and cellular characteristics of the filamentous fungal isolates grown for 7 days on SDA. Photomicrographs were taken at 100X (middle column) and 400X (right column).

The colonies of Isolate 2 were white-colored, cottony-like at the early stage, and had pin-like sporangia and sporangiophores. They had no distinct stolon structure, instead the sporangiophore produced branches that were aseptate. These are characteristics of the genus *Mucor* under the Phylum Oomycota (Figure 2 second row). On the other hand, isolate 3 colonies were deep green in color, cottony-like at early stage, and had branched conidiophores. There were no stolons and septae, and the spores were not enclosed in a sporangium, which characterize the genus *Trichoderma* (Figure 2 third row).

In a study done by Manguilimotan and Bitacura (2018), eight Cd-tolerant filamentous fungi were isolated from the coastal waters and sediments near the effluent sites of the industrial plants in Barangay Ibo, Lapu-Lapu City, Cebu. In terms of Cd biosorption activity, only three isolates were found to be most efficient. Two of them belonged to genus *Aspergillus* and one to genus *Penicillium*. Neither of those two genera was isolated in the present study. Furthermore, *Rhizopus* and *Trichoderma* were also among the filamentous fungi isolated by Zafar et al (2007) from agricultural soils with long term input of wastewater from their municipality and industry.

Heavy Metal Tolerance of the Fungal Isolates

The fungal isolates were screened for their tolerance to 1mM each of Cd, Cu, Fe and Zn (Figure 3). Figure 4 shows that Cu was the most tolerated heavy metal by *Rhizopus* sp. followed by Fe, Zn, and Cd. *Mucor* sp., was most tolerant to Fe and Zn followed by Cu then Cd. *Trichoderma* sp. was most tolerant to Fe then Zn and Cu, but least tolerant to Cd. Figure 5 shows that *Rhizopus* sp. exhibited the highest tolerance index to all the heavy metals tested compared to that of *Trichoderma* sp. and *Mucor* sp.

In a similar study conducted by Zafar (2007), *Rhizopus* and *Trichoderma*, isolated from heavy metal contaminated agricultural soils, were found to be tolerant to Cd, Ni, Cr, Cu, and Co. Meanwhile, Oladipo et al (2018) found *Rhizopus* and *Trichoderma*, isolated from gold and gemstone mine site soils, as tolerant to Cu, Pb, and Fe.



Figure 3. Growth of the fungal isolates after 7 days on PDA with and without 1mM heavy metal treatment

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Figure 3 continued



Figure 4. Tolerance indices of fungal isolates (Mean \pm SD) to 1mM of heavy metal salts. Levels having the same letter designations are not significantly different at p<0.01



Figure 5. Comparative tolerance indices (Mean \pm SD) of the fungal isolates to 1mM heavy metal salts. Levels having the same letter designations are not significantly different at p<0.01

Minimum Inhibitory Concentration of the Heavy Metals Against Rhizopus sp.

The minimum inhibitory concentration of the four heavy metals against the fungal isolate is summarized in Table 2 and Figure 6. *Rhizopus* sp. was shown to tolerate Cu concentrations of 10 to 15mM only. Cadmium, on the other hand, was the most growth-inhibiting to the fungal isolate. Maximum Cd concentration in which the fungus was able to grow was between 5 to 6.5mM. The highest heavy metal concentration that *Rhizopus* sp. can grow is in Fe with a minimum inhibitory concentration of 30mM<MIC≤35Mm, followed by Zn with 25mM<MIC≤30mM.

Table 2. Range of average Minimum Inhibitory Concentration (MIC) of the heavy metals tested with *Rhizopus* sp.

Fungal Isolate	Minimum Inhibitory Concentration (mM)			
	Cd ²⁺	Cu ²⁺	Fe ²⁺	Zn ²⁺
Rhizopus sp.	5 <mic≤6.5< td=""><td>10<mic≤15< td=""><td>30<mic≤35< td=""><td>25<mic≤30< td=""></mic≤30<></td></mic≤35<></td></mic≤15<></td></mic≤6.5<>	10 <mic≤15< td=""><td>30<mic≤35< td=""><td>25<mic≤30< td=""></mic≤30<></td></mic≤35<></td></mic≤15<>	30 <mic≤35< td=""><td>25<mic≤30< td=""></mic≤30<></td></mic≤35<>	25 <mic≤30< td=""></mic≤30<>

The growth of the fungal isolates observed in this study was similar to the report of Ezzouhri et al (2009) wherein the growth pattern first exhibited a lag phase, then retardation phase, after which no additional enhancement in growth was observed. In the present study, there was a longer lag phase (adjustment period) in the heavy metal treatments compared to the control. As the concentration of the heavy metal was increased, the colony extension decreased (Figure 6). This may mean that the fungus developed tolerance or adaptation during incubation as suggested by Ezzouhri et al (2009). Furthermore, increasing the heavy metal concentration appeared to slow down the growth of the mycelia. The presence of a longer lag phase at higher heavy metal concentrations suggests that tolerance development or adaptation may have occurred during incubation. According to Gadd (1993), a reduction in the growth of a fungus is a natural response to toxicants but an increase in the length of the lag phase is not always present. Additionally, dependency on lag time is not always present according to the study of Darlington and Rauser (1988) wherein *Penicillium involutus* did experience an extended lag phase. Lengthening of lag phase is not always present probably because the fungus is tolerant enough to higher concentrations of the heavy metals.



Figure 6. Growth of *Rhizopus* sp. after 7 days on PDA with different concentrations of Cd, Cu, Fe and Zn salts



Figure 6 continued

Among the fungal isolates, *Rhizopus* sp was the most tolerant to Zn and Fe (Figure 4). Its colony color became lighter and the mycelia grew thinner compared to the control. These were manifestations of the stress caused by the heavy metals. Cu is a co-factor in many enzymatic processes and is the 3^{rd} most abundant transition heavy metal found in living organisms (Brandolini et al 2002). *Rhizopus* sp. showed high tolerance to 1mM of Cu, but when the concentration was increased its growth was slow. It was the third heavy metal that the fungus could tolerate the most. The fungus grew smaller compared to those that grew on Fe and Zn. There was blue coloration of the mycelium inoculated on the Cu-treated medium, which may indicate that Cu ions were bound to the fungal cell walls. Anand et al (2006) and Ezzouhri et al (2009) also reported similar observations. The MIC of *Rhizopus* sp. to Cu was $10 < MIC \le 15mM$ which was lower than that of *Aspergillus* sp. which was 15-20mM as reported by Ezzourhi et al (2009).

Cadmium is a by-product of Zn production and is a well-known carcinogen and teratogen (New Jersey-DOH 2007). In Figure 3, the growth of the fungus in Cd-treated medium was very low compared to those cultured with other metals, making it the most toxic heavy metal compared to Cu, Fe, and Zn. In a similar study by Ahmad et al (2005), *Rhizopus* sp. isolated from wastewater-treated soil had an MIC of 1μ g mL⁻¹. Additionally, it was found to be more biosorptive than *Aspergillus* sp. The low MIC may have been due to the high toxicity of the heavy metal.

Tolerance of filamentous fungus may also be directly related to the concentration of the heavy metals in the sediments (Ahmad et al 2005). In the sediment samples collected from the settling pond, the concentrations of Fe and Zn were much higher than those of Cu and Cd. This heavy metal tolerance is very important because when there is no tolerance to heavy metals, they grow slow and eventually die. It would be cost ineffective if they are used in contaminated areas for bioremediation.

In this study, the MIC of *Rhizopus* sp. to Cd appeared to indicate that its tolerance to this metal was not fully developed which may be due to the low concentration of Cd in the settling pond (Table 1). For Cu, its concentration in the settling pond was higher, which could have induced the fungus to develop more

tolerance to the metal. *Rhizopus* sp. was most tolerant to Cu at 1mM or 159.609mg kg⁻¹ but tolerance decreased at much higher concentrations of Cu. In contrast, high tolerance of *Rhizopus* sp. to Fe and Zn was still observed even at much higher concentrations. This means that the fungus may have developed higher tolerance to the heavy metals in response to the high metal concentrations in the pond sediments (Table 1). This may be an indication that *Rhizopus* sp. has a high bioremediating potential in heavy metal polluted waters.

In a study done by Pal et al (2010), a heavy metal tolerant strain of *Rhizopus arrhizus* accumulated lead in the cell walls and that cellular metabolism could be involved in the bioaccumulation of the heavy metal by the fungus. Moreover, Tsekova and Galabova (2003) reported that the acid phosphatase production in *Rhizopus delemar* in and out of the cells after exposure to Cu ions is necessary for its heavy metal resistance and bioaccumulation.

CONCLUSION

Three filamentous fungi were successfully isolated from the sediments and were identified as *Rhizopus* sp., *Mucor* sp. and *Trichoderma* sp. The most tolerant isolate to all four heavy metals tested was *Rhizopus* sp. It tolerated Cu the most, and Cd the least. The MIC values of Cd, Cu, Fe, and Zn to *Rhizopus* sp. were at 5mM<MIC≤6.5mM, 10mM<MIC≤15mM, 30mM<MIC≤35mM and 25mM<MIC≤30mM, respectively. These results indicate that *Rhizopus* sp. is a potential candidate species for bioremediation of Cu, Fe, and Zn-contaminated waters. It is recommended that further studies on its heavy metal biosorption, bioaccumulation capacity, and other mechanisms of bioremediation be conducted.

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